APHE-1 AND APHE-2, TWO NEW ANTIMICROBIAL AND CYTOTOXIC ANTIBIOTICS

II. PHYSICO-CHEMICAL PROPERTIES AND STRUCTURE ELUCIDATION

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Two new pyrazolo-isoquinolinone antibiotics, APHE-1 and APHE-2, have been isolated from
the culture filtrate and mycelia of *Streptoverticillium griseocarneum* NCIMB 40447. Molecular formulae were established as $C_{13}H_{12}N_2O$ for APHE-1 and $C_{14}H_{14}N_2O$ for APHE-2, by elemental analysis, NMR and mass spectra. 2D NMR techniques $(^1H-^{1}H$ COSY-45 and $^1H-^{13}C$ correlated spectroscopy) have been applied to establish their structures. \mathbf{r} is establish the best applied to establish the structures.

In the preceding paper we have described the fermentation, isolation and biological activities of APHE-1 and APHE-2, two new antimicrobial and cytocidal antibiotics¹⁾. In this paper, we report the physico-chemical properties and structural elucidation of APHE-1 and APHE-2. physico-chemical properties and structural elucidation of APHE- 1 and APHE-2.

Physico-chemical Properties

The antibiotics APHE-1 and APHE-2 were isolated as yellowish powders. They were easily soluble
in methanol, ethanol, acetone, ethyl acetate and chloroform, slightly soluble in benzene but insoluble in hexane, petroleum ether and water. Both compounds gave negative ninhydrin reaction. The molecular hexane, petroleum ether and water. Both compoundsgave negative ninhydrin reaction. The molecular formulae were determined to be $C_{13}H_{12}N_2O$ for APHE-1 and $C_{14}H_{14}N_2O$ for APHE-2 by EI-M and elemental analysis. Additional physico-chemical and chromatographic properties are given in Table1.

^a Silica gel TLC (Merck); solvent: chloroform - methanol (97:3). Silica gel TLC(Merck); solvent: chloroform- methanol (97 : 3).

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Structural Determination
The ion $m/z = 197$ was present in both MS spectra (Table 1) and was orginated from M⁺ peak (base peak) by the loss of a fragment of 15 (CH₃) and 29 (C₂H₅) mass units for APHE-1 and APHE-2, respectively. The fragmentation sequence from this ion led to the same essential fragment ions, suggesting respectively. The fragmentation sequence from this ion led to the same essential fragment ions, suggesting the presence of the same or a similar $(C_{12}H_9N_2O)$ structural unit. The UV spectra (Table 1) we identical for both compounds and exhibited characteristic absorption maxima at 265, 280 and 296nm, indicating a similar aromatic chromophore. Their IR spectra (Table 1) showed an amide carbonyl absorption at 1640 and 1579 cm⁻¹ for APHE-1 and 1635 and 1575 cm⁻¹ for APHE-2 as well as a NH band.

The ¹H and ¹³C NMR spectra in CDCl₃ solutions exhibited very simple signals in the upfield region which were attributed to an ethyl and a *n*-propyl group for APHE-1 and APHE-2, respectively, in agreement with MS spectra. The remaining signals of the two compounds, due to a highly unsaturated $(C_{11}H_7N_2O)$ moiety, appeared at low field and showed similar chemical shifts and multiplicities. The wide singlet at ca. δ 8.5 in the ¹H NMR spectra (shifted at δ 10.63 for APHE-2 in acetone- d_6 solution), exchanged with D₂O and was attributed to NH proton. The signal at ca. δ 163 in the ¹³C NMR spectra confirmed the amide carbonyl group. Between δ 7.1 ~ 7.9 there were six signals for methine groups which were confirmed by the corresponding carbon resonances in the 13 C NMR spectra. The spectra also were confirmed by the corresponding carbon resonances in the 13C NMRs per sonal α nm α in the spectra also be spectra. The spectra also be spectra also be spectra. The spectra also be spectra also be spectra also be exhibited four quaternary carbon signals in the region σ 105 \sim 148. These facts suggested the same

features for this structural unit and agreed with an heteroaromatic system.

In order to propose a plausible structure for these compounds the ¹H and ¹³C NMR spectra of APHE-2 were analyzed in depth by application of 2D NMR techniques^{2,3)} and double resonance (DR) A HE-2 were analyzed in depth by application of 2D NMR techniques³ and double resonance (DR) experiments. In particular, COSY-454) and heteronuclear *H-13C correlated spectroscopy (XHCORD)5'6) were used.
The COSY cross-connectivity patterns (Fig. 1) revealed that the doublet at δ 7.53 only exhibited a

large correlation with the NH signal, and no correlations were found for the slightly broadened singlet at δ 7.19. A weak cross-peak at δ 7.82/7.44 and two strong cross-peaks at δ 7.82/7.25 and 7.44/7.25 were observed. To strengthen these facts, DR experiments were performed. Thus, the saturation of the multiplet at δ 7.82 simplified the signal at δ 7.44 into a doublet of doublets with splitting of ca. 7.0 and 1.5 Hz. This experiment also showed a loss of a coupling of approximately 1.5 Hz at the signal centered at δ 7.29 which appeared as an apparent triplet with a splitting of ca. 7.0 Hz. Further interpretation of DR 8 7.29 which appeared as an apparent triplet with a splitting of ca. 7.0Hz. Further interpretation of DR experiments, in which the signals at σ $/44$, $/35$ and 8.44 were saturated, committed the above considerations. The study of the ¹H and COSY-45 spectra and DR experiments in acetone- d_6 led to the same results.
The analysis of the respective signals allowed the establishment of the proton magnetic parameters

of APHE-1 and APHE-2 collected in Table 2. On the basis of these statements, the four-spin ABMX system at δ 7.25, 7.29, 7.44 and 7.82 was attributed to a 1,2-disubstituted phenyl ring. The value of the coupling constants between the CH proton at 7.53 and the NH proton (2.6 Hz) was similar to those found in five-membered heteroaromatic systems^{7,8)} and is compatible with the presence of a substituted diazole. Moreover, both rings might be linked by the carbonyl amide and the isolated CH $(\delta 7.19)$ groups.

 13 C chemical shifts are tabulated with signal multiplicity, obtained from DEPT experiments, in Table 2. Once the resonances of the respective protons were established, the analysis of the ${}^{1}H^{-1}{}^{3}C$ correlated 2. Once the resonances of the respective protons were established, the analysis of the 1H-13Ccorrelated s_{p} is the spectrum of the identification and assignment of the following signals: $\frac{1}{2}$

| ¹ H δ (ppm) ^a | | | J_{HH} (Hz) ^b | | | ¹³ C δ (ppm) ^c | | |
|--|---------|----------|----------------------------|--------|--------|---|---------------------|---------------------|
| Assignment | APHE-1 | $APHE-2$ | Assignment | APHE-1 | APHE-2 | Assignment | APHE-1 | APHE-2 |
| $1-H$, br s | 8.53 | 8.44 | $1-H \sim 2-H$ | 2.6 | 2.6 | $C-2$, d | 121.53 | 121.46 |
| $2-H, d$ | 7.53 | 7.53 | $5-H \sim 6-H$ | 7.4 | 7.6 | $C-3. s$ | 105.82 | 106.00 |
| $4-H, s$ | 7.17 | 7.19 | $5-H \sim 7-H$ | 1.4 | 1.4 | $C-3a$, s | 147.21 | 147.00 |
| $5-H$, m | 7.44 | 7.44 | $5-H \sim 8 \text{ H}$ | 0.8 | 0.8 | $C-4$, d | 119.36 ^d | 119.72 ^d |
| $6-H, m$ | 7.31 | 7.29 | $6 - H \sim 7 - H$ | 7.1 | 7.1 | $C-4a$, s | 136.14 | 136.18 |
| $7-H$, m | 7.23 | 7.25 | $6-H \sim 8-H$ | 1.5 | 1.6 | $C-5$, d | 111.46 | 111.44 |
| $8-H, m$ | 7.83 | 7.82 | $7-H \sim 8-H$ | 7.3 | 7.4 | $C-6, d$ | 122.94 | 122.90 |
| $1'$ -H | 2.90, q | 2.87, t | $1'$ -H \sim 2′-H | 7.5 | 7.4 | $C-7$, d | 120.79 | 120.75 |
| $2'$ -H | 1.42. t | 1.90, m | $2'$ -H \sim 3'-H | 7.5 | 7,4 | $C-8$, d | 119.87 ^d | 119.92 ^d |
| $3'$ -H, t | | 1.06 | | | | $C-8a$, s | 124.00 | 124.06 |
| | | | | | | $C-9$, s | 163.69 | 163.40 |
| | | | | | | $C-1$ ', t | 21.62 | 30.70 |
| | | | | | | $C-2'$ | 11.22, a | 20.58, t |
| | | | | | | $C-3'$, q | | 13.70 |

Table 2. ¹H and ¹³C NMR data for APHE-1 and APHE-2 (299.949/75.429 MHz, CDCl₃).

 $^{\circ}$ Error \pm 0.1 Hz.

 $\frac{1}{\sigma}$ Values may be interchanged d Values may be interchanged.

13.70/1.06 (CH₃), 20.58/1.90 (CH₂), 30.70/2.87 (CH₂), 120.75/7.25; 122.90/7.29; 111.44/7.44; 119.92/ 7.82 (CH, 1,2-disubstituted phenyl ring), 119.72/ 7.19 (isolated CH) and 121.46/7.53 (CH-NH, diazole system). The signals at 119.72 and 119.92 could not be distinguished and the correlation with
the proton signal at δ 7.19 was very weak.

 \mathbf{r} , \mathbf{r} , \mathbf{r} was very weak.

Fig. 2. Structures of APHE-1 and APHE-2.

 μ ₁₄₇. The spectroscopic studies have astablished that these sampaumds are sendered betapticularly 40447. The spectroscopic studies have established that these compounds are condensed heterotricycles which only differ in the nature of an aliphatic chain; an ethyl and a n -propyl group for APHE-1 and APHE-2, respectively. Taking into account the spectroscopic data as well as substituent steric and electronic effects on proton^{7,8)} and ¹³C chemical shifts^{8,9)} and the magnitude of the ¹H-¹H coupling constants^{7,8)}, a b-fused 1(2H)-isoquinolinone system is proposed as the most probable structure. These antibiotics are considered as 3-substituted $1H$ -pyrazolo $[2,3-b]$ isoquinolin-9-ones (Fig. 2). This structure data. The assignment of the respective proton and carbon signals for the proposed structure are shown $\frac{1}{2}$ as signals for the respective proton and carbon signals for the proposed structure are shown in Table 2.

Experimental

The melting points were uncorrected. The IR spectra were recorded on KBr pellets on a Perkin-Elmer 883 spectrophotometer. UV spectra (λ_{max} in nm) were recorded in MeOH solutions on a Beckman DU-50 spectrometer. The mass spectra were carried out in a Hewlett-Packard 5987A spectrophotometer by EI. The elemental analysis were performed on a Perkin-Elmer 240B elemental analyzer. The ¹H, ¹³C and 2D NMR spectra as well as double resonance (DR) and DEPT experiments were recorded on a Varian UNITY-300-FT spectrometer (299.949 MHz for ¹H and 75.429 MHz for ¹³C), using solutions ca. 0.02 M in CDC₁ and ecotons d , with TMS as the internal reference at $2029V$ in CDC13 and acetone-^ with TMSas the internal reference, at 293°K.

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